
Structural bioinformatics

EasyVS: a user friendly web based tool for molecule library selection and structure-based virtual screening

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Abstract

Summary: EasyVS is a web-based platform built to simplify molecule library selection and virtual screening. With an intuitive interface, the tool allows users to go from selecting a protein target with a known structure and tailoring a purchasable molecule library to performing and visualising docking in a few clicks. Our system also allows users to filter screening libraries based on molecule properties, cluster molecules by similarity and personalise docking parameters.

Availability and implementation: EasyVS is freely available as an easy-to-use web interface at <http://biosig.unimelb.edu.au/easyvs>

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Supplementary information: Supplementary data are available at *Bioinformatics* online.

1 Introduction

Structure-based virtual screening has been widely and successfully used in early stages of drug development, aiding in the identification of potential hits and guiding further experimental validation (Cheng *et al.*, 2012). Molecular docking is one of the most widely used virtual screening approaches, which uses the three-dimensional structure of a target protein to predict the predominant binding mode of a small molecule with the target of interest. In this way, docking can be used to evaluate a large library of molecules and identify those most likely to interact with the

target in the desired manner. This has been a powerful tool in the identification of initial hits, significantly reducing the chemical space to experimentally test, and increasing the proportion of positive compounds being screened.

Significant improvements in docking protocols (Di Muzio *et al.*, 2017), scoring functions (Pires *et al.*, 2016), and molecule libraries (Sterling *et al.*, 2015), and the greater availability of computational power, has made virtual screening a more tractable and reliable hit identification strategy. Despite this, current virtual screening approaches typically require specialist computational and technical expertise.

In order to make virtual screening more friendly and accessible to a wider audience, we propose EasyVS (<http://biosig.unimelb.edu.au/easyvs>), a

web-based, efficient and intuitive system that allows users to go from defining a protein structure and molecule library to performing docking in a few clicks. Our system allows users to optimize their screening library based on their properties (and define a chemical space of interest). Through a molecule clustering approach, we can more rapidly screen a larger chemical space, and present the top solutions to the user.

2 Platform description

Initially users are asked to define their target of interest by either uploading a structure of interest, or using the biological assembly of a previous experimental structure deposited in the RCSB by providing the PDB accession code.

A

1. Molecules Selection

Click to Upload SDF file

OR

Choose one or more molecule databases:

- Select/DeSelect all
- Drugbank (7411 Ligands)
- HMDB (12095 Ligands)
- Maybridge (5772 Ligands)
- CHEMBL (1370423 Ligands)
- Supernatural (222893 Ligands)
- Zinc purchasable (1322996 Ligands)
- chembridge_core (79502 Ligands)
- chembridge_express (504034 Ligands)

2. Filter by property

Pre-selections: Custom, Set to RO5, Set to RO3

Properties:

Atoms: 1 to 600

Molecular weight: 0 to 200

Hydrogen donors: 0 to 100

Hydrogen acceptors: 0 to 100

Rings: 0 to 20

Rotatable bonds: 0 to 100

LogP: -20 to 20

3. Filter by similarity

Similarity: 0.85

Filter molecules

Summary

After filter: 7943 ligands

Time estimation: 66.2 hours

B

Docking data

Show 5 entries

| Molecule depiction | Mol. Identifier | Affinity (kcal/mol) | Affinity (pMolar) | View poses | Arpeggio | pkCSM | Atoms | Mol. Weight | Hydrogen donors | Hydrogen acceptors | Rings | LogP |
|--------------------|-----------------|---------------------|-------------------|----------------------|---------------------|---------------------|-------|-------------|-----------------|--------------------|-------|------|
| | DB08703 | -13.80 | 0.76 | View | Run | Run | 32 | 424.50 | 1 | 4 | 6 | 4.91 |
| | DB087113 | -13.30 | 3.49 | View | Run | Run | 34 | 469.49 | 4 | 5 | 4 | 3.31 |
| | CHEMBL297792 | -13.00 | 5.18 | View | Run | Run | 32 | 464.57 | 3 | 4 | 4 | 3.31 |
| | CHEMBL52171 | -12.35 | 4.34 | View | Run | Run | 23 | 311.16 | 2 | 5 | 3 | 3.09 |
| | CHEMBL273264 | -12.70 | 5.49 | View | Run | Run | 26 | 347.38 | 5 | 4 | 3 | 2.85 |

Showing 1 to 5 of 6,675 entries

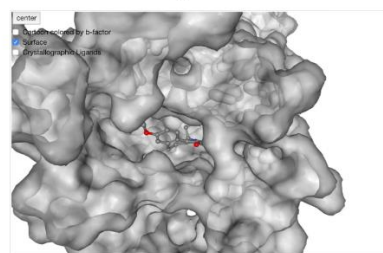
Previous 1 2 3 4 5 ... 1335 Next

Visualisation

Protein 4IAR was docked to DB08703 with Affinity of 0.76nM

Download the PDB file of target

Download all poses in one SDF file



Click to show/hide docking results

- ☒ Pose 1: -13.80 kcal/mol [SDF file](#)
- ☐ Pose 2: -12.80 kcal/mol [SDF file](#)
- ☐ Pose 3: -11.50 kcal/mol [SDF file](#)
- ☐ Pose 4: -11.50 kcal/mol [SDF file](#)
- ☐ Pose 5: -11.30 kcal/mol [SDF file](#)
- ☐ Pose 6: -11.00 kcal/mol [SDF file](#)
- ☐ Pose 7: -10.70 kcal/mol [SDF file](#)
- ☐ Pose 8: -9.60 kcal/mol [SDF file](#)
- ☐ Pose 9: -8.90 kcal/mol [SDF file](#)
- ☐ Pose 10: -7.90 kcal/mol [SDF file](#)

Fig. 1. EasyVS web interface. Once a protein target is selected, (A) users are prompted to select a molecule library from available databases and property filters. (B) Docking will be performed on the selected target/library set and best poses shown as an interactive molecule visualization.

The selected structure is then analyzed using Ghecom (Kawabata, 2010) to identify druggable pockets. While by default the largest pocket is chosen for docking, users may select another pocket of interest for screening, and can refine the boundaries and docking parameters used (Fig. S1 of Supplementary Materials). These parameters include box size

and position (which can be set to any of the identified pockets and finely adjusted by the user) and depth of the search.

In the next step, users are asked to define the set of molecules to be assessed via docking. EasyVS currently supports the small molecule databases ChEMBL 24_1 (Gaulton *et al.*, 2017), HMDB 4.0 (Wishart *et al.*, 2018), Drugbank 5.0 (Wishart *et al.*, 2018), Maybridge¹, Super Natural II (Banerjee *et al.*, 2015), Chembridge (Desai *et al.*, 2004) and Zinc15 (Sterling *et al.*, 2015), which together comprise over 230 million molecules.

There are many filters available for refining these molecular libraries, including by molecular weight, number of acceptors or donors of hydrogen, *logP* (or only selecting Lipinski's Ro5 molecules), fragments or natural products (Figure 1A). Once the molecule library has been selected, users have the option to cluster molecules by similarity to improve screening performance. If users opt to perform clustering, one representative molecule from each group is randomly selected for docking. Users have the option to select the level of similarity used during clustering, which will change the number of clusters and, therefore, the number of molecules that will proceed for docking stages.

Molecule docking of the selected compound library is then performed using Autodock Vina (Trott *et al.*, 2010). After each molecule is docked, the NNScore function (Durrant, *et al.*, 2011) is used to estimate the binding affinity (K_D) and binding free energy of the docked poses using a trained neural network, in order to evaluate the quality of the poses generated by Autodock Vina. Users can also rapidly analyse the intramolecular interactions using Arpeggio (Jubb *et al.*, 2016) and predict pharmacokinetic properties of top hits using pkCSM (Pires *et al.*, 2015). While the docking is running, users can view the results in real time to analyze best poses of selected molecules (Figure 1B) as well as include additional molecules for docking. Further exploration of the chemical space of top docked ligands is available by an additional round of virtual screening, using compounds that are structurally similar to the ligand of interest.

The EasyVS docking protocol was validated using two different benchmarks. We performed a redocking procedure for a selection of eight GPCR-ligands complexes with available crystallographic structures. Ligands have been successfully redocked with an average RMSD of 0.98 Å (Table S1 of Supplementary Materials). We have also created decoy libraries using DUD-e (Mysinger *et al.*, 2012) for the same set of proteins considered for redocking. The docking scores for real ligands were considerably higher than those obtained for the decoys (p-value < 0.001, Table S2 of Supplementary Materials), demonstrating the robustness of the EasyVS docking protocol.

3 Conclusions

Here we present EasyVS, a freely available, user-friendly platform for simplifying molecule library construction and docking. EasyVS allows users to choose molecules from well-established and diverse databases, including fragments, approved drugs and natural products, and perform the docking with just a few clicks. We also show EasyVS was successful in identifying GPCR ligands (Supplementary Materials) as a case study. We believe this will be an invaluable tool for the exploratory stages of hit identification, allowing for the selection either stringent or very diverse sets of molecules for virtual screening and the intelligent assessment of different small molecule chemical spaces.

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Conflict of Interest: none declared.

¹ <https://www.maybridge.com>

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